TITLE PAGE

Residues of Imidacloprid and Imidacloprid Metabolites in Nectar, Blossoms, Pollen and Honey Bees Sampled from a French Summer Rape Field and Effects of These Residues on Foraging Honeybees

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STATEMENT OF COMPLIANCE

This study was conducted in compliance with the Principles of Good Laboratory Practice (Chemicals Law (ChemG) of July 25, 1994, Annex 1 and OECD Principles of Good Laboratory Practice (GLP) of November 26, 1997 [C(97) 186/Final]).

Signature:

[redacted]

Study Director

Title Date

Responsible Analyst

Title Date

CERTIFICATION OF AUTHENTICITY

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Title

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1.0 SUMMARY

Report: [redacted] (1999): Residues of Imidacloprid and Imidacloprid Metabolites in Nectar, Blossoms, Pollen and Honey Bees Sampled from a French Summer Rape Field and Effects of These Residues on Foraging Honeybees
Bayer AG, unpublished report No: SXR/Am 001; 1999/04/30.
(Appendix I reports the data from study RA-2042/98)

Guidelines: Internal Testing Method
Deviations: not applicable

GLP: yes (certified laboratory)

Material and methods: Poncho FS 500, a.i. content: 78.3 g/L Beta-Cyfluthrin & 428.2 g/l Imidacloprid; specification (formulation No.: 030 based on 06200/0029, developmental No.: 00195939); under field conditions small beehives (appr. 5,000 honeybees) were caged on flowering summer rape plots (drilling rate: 5 kg/ha) as a sampling device for rape nectar and rape pollen. Nectar was also directly sampled from flowers via micropipettes. In addition, flowers were sampled by hand. The honeybees used as sample collectors were observed for signs of behavioral impacts. All samples including the honexbees were subjected to a residue analysis for imidacloprid and its relevant metabolites.

Dates of biological work: June 15 - 18, 1998
Dates of analytical work: June 30 – July 22, 1998.

Findings: Residues in rape plant matrices and in the foraging honeybees

Type of Sample	Residue Level [mg/kg] *		
	Imidacloprid	Olefin-NTN	Hydroxy-NTN
Control Samples			
Honeybees before exposure	< 0.01	< 0.01	< 0.01
Rape nectar sampled by bees	--	--	--
Rape nectar sampled with micro-capillaries from the flowers	< 0.01	< 0.01	< 0.01
Rape blossoms	< 0.01	< 0.01	< 0.01
Rape pollen sampled by bees	--	--	--
Treatment Samples			
Honeybees before exposure	< 0.01	< 0.01	< 0.01
Rape nectar sampled by bees	< 0.01	< 0.01	< 0.01
Rape nectar sampled with micro-capillaries from the flowers	< 0.01	< 0.01	< 0.01
Rape blossoms	< 0.01	< 0.01	< 0.01
Rape pollen sampled by bees	< 0.01	< 0.01	< 0.01

* Limit of quantitation: 0.01 mg/kg

Observations: No behavioral impacts (e.g. apathy, exaggerated motility, discoordinated movements) or suspicious mortality was observed on the honeybees used for collecting rape nectar and rape pollen. At the time of sampling, aphids were observed on the rape plants.

2.0 INTRODUCTION

According to EU directive 91/414/EWG the impacts of pesticides on honeybees have to be examined. Besides the intrinsic toxicity of a pesticide the concentration to which a honeybee may be exposed under field conditions is an integral component for the hazard assessment. The present study aims to examine the exposure in greater detail for a refined risk assessment.
The rape samples were analysed for residues of imidacloprid and its olefin- and hydroxy metabolites. These metabolites were considered as relevant, since they have a chemical structure closely related to the parent molecule and were observed in plant metabolism studies in significant proportions (up to approx. 10 %).

3.0 EXPERIMENTAL

3.1 Test Substance

Test Substance:	Poncho FS 500
Active Ingredient(s):	(a) Beta-Cyfluthrin (FCR 4545) (b) Imidacloprid (NTN 33893)
Chemical Name(s) of AI(s):	(a) CYCLOPROPANECARBOLXYLIC ACID,-= 3-(2,2-DICHLOROETHENYL)-2,2-= DIMETHYL-,CYANO(4-FLUORO-3-= PHENOXYPHENYL)METHYL ESTER (b) 1-<(6-CHLORO-3-PYRIDINYL)-METHYL>-= N-NITRO-2-=IMIDAZOLIDINIMINE
CAS Number of AI(s):	(a) 68359 - 37 - 5 (b) 138261 - 41 - 3
Indikation:	Seed dressing
Product Number	0195939
Formulation/Batch Number:	FL 0030 based on form. no. 06200/0029
No. of Certificate:	FAR-No. 446-01
AI Content (acc. to Analysis):	(a) 78.3 g/l (b) 428.2 g/l
Analytical Method:	(a) GLC, int. Std. (b) HPLC, ext. Std.
Date of Analysis:	February 4, 1998
Expiry Date:	February 4, 1999
Physical Appearance:	dark blue suspension
Specific Density:	1.151 g/ml
Storage Conditions:	Room temperature
Seed Dressing Rate(s) Tested in the Study:	2.5 l Poncho 500 FS per 100 kg oilseed rape (= 1050 g/dt Imidacloprid & 200 g/dt beta-Cyfluthrin) (analytical findings *: 1045 g/dt Imidacloprid).
Seed Drilling Rate Tested in the Study:	5 kg seed per ha
Safety Precaution:	Routine hygienic precautions

3.2 Reference Substance

For this type of material and use pattern, a reference compound is not specified.

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* Dressed seeds were analysed for imidacloprid only.

3.3 *Execution of the Test*

The sampled study plots were drilled on March 19, 1998 (reserve plots drilled on April 14, 1998). Sampling of nectar, flowers and honeybees and the behavioral observations were performed between June 15 and 18, 1998.

Sponsor:	BAYER AG GB Plant Protection Marketing - Seed Treatment [redacted] D-40789 Monheim
Study Director:	[redacted]
Cultivar Manager:	
Trials Officer:	
Responsible Analyst:	
Study Technicians:	
Quality Assurance:	H. Grigat / A. Stollenwerk
Laboratory Study Number:	SXR/Am 001

3.4 *Origin of Honeybees*

The honeybees used for pollen and nectar sampling were supplied by a commercial French beekeeper ([redacted] Before use, the beehives stood at a forested area about 10 km apart from the trial site. The beehives used for the test were transported to the study site in the evening of June 16 and returned to the original place on June 18, 1998.

3.5 *Procedure of Seed Dressing and Dispatch of Test Product*

Rape seeds (variety: „Lisonne"; TGW: 2.8 g) were coated in a Centauer coating machine. Some 562.5 ml Poncho FS 500 were added to 22.5 kg rape seed together with 450 g Talcum blue (Product no. 868426) and mixed over 45 seconds at 300 RPM.

3.6 *Location of the Trial Site and Description of the Study Plots*

The trial site was located in the vicinity of Conches between la Neuve Lyre and la Vieille Lyre in Northern France.

There were two rows of 4 rape planted plots each on the trial site (Fig. 1). Each plot had a size of 4x20 m with 1 m space between adjacent plots and 8 m space between the two plot rows. Each plot was staked out with marking sticks prior to the beginning of the experimental part.

3.7 *Drilling of the Rape Seed and Calibration of the Seed Machinery*

The control plot (no. 1 in Fig. 1) was drilled with 5 kg/ha untreated rape seed whereas the treatment plot (no. 2 in Fig. 1) received 5 kg/ha rape seed coated with 2.5 l/ha Pocho FS 500.

Prior to sowing the proper functioning of the equipment was tested. This machinery control is documented. The equipment was adjusted according to the preconditions (e.g. seed density).

3.8 Cultivation of the Plots

Treated and untreated plots were cultivated in the same way according to the practice of the region. Before initiation of sampling, no protection treatments other than the seed treatment was necessary.

3.9 Sampling Procedure

Installment of bee hives

At the time of full rape blossom, tents of 4x4 m and 2 m height were installed on the control and the treatment plot (see Fig. 1). The tents consisted of an aluminium frame covered by gauze material (2x2 mm mesh size). For handling purposes, a walkway was created by removing all plants along a 50 cm transect between the tent entrance and the opposite end. One bee hive was placed at the end of the walkway opposite to the entrance in the treatment plot. In the control plot, no bee hive could be installed since the prepared beehive was not in a condition which permitted a transport. In the treatment plot, the hive entrances was disclosed and honeybees were allowed to forage on the study plot within the tent area. Before placing the beehives on the treatment plots, appr. 100-200 honeybees were sampled to get blank samples of honeybees and honeybulbs for the residue work. The sampled honeybees were processed as described in the subsequent chapter.

Sampling of Nectar from the Honeybulbs

On the day of hive installment and the following two days a total of about 200 honeybees were sampled with tweezers directly from rape flowers after watching them for feeding over about 10-30 seconds. All sampled bees were killed by freezing (dry ice). Dead bees were stored on dry ice in the field and, at the end of each sampling day at the latest, transferred to a refrigerator (-20°C). At the end of the study the samples were shipped to Monheim on dry ice and further retained at 20°C until preparation of the honeybulbs. Honeybulbs were prepared by cutting the frozen bees into halves between the thorax and the abdomen and removing the filled honeybulbs by tweezers. All honeybulbs from one treatment group were pooled witin an Eppendorf cap which was stored on dry ice. After all honeybees of a respective treatment were prepared (at the end of each preparation day at the latest), sampled honeybulbs were stored in a refrigerator at -20°C until residue analysis (see 3.10).

Sampling of Pollen from the Honeybees

From the prepared bees, pollen pockets were removed and stored in a refrigerator at -20°C until residue analysis (see 3.10). From the blank samples, not sufficient pollen could be obtained for residue analysis.

Sampling of Nectar from the Rape Flowers

The day before sampling, between 10 and 20 flowering rape plants outside the caged area were covered by plastic bags to prevent insects from foraging on that flowers. On the next day, nectar was directly drawn from that flowers by 5 µl micropipettes After sampling, the micropipettes were emptied into a 1.5 ml Eppendorf tube which was stored on dry ice in the field. At the end of each sampling day at the latest, these samples were transferred into a refrigerator (-20°C) where they were retained until residue analysis (see 3.10).

As a check of possible contamination of the sampled material from e.g. soil particles, dust, sampling people had to fill a second Eppendorf tube with uncontaminated tap water

during sampling activities. Filling of this checking tube was done in several steps (5-10 pipetting events) from a water storage bottle. The analysis of these check samples revealed no residues indicating that the sampling people did not transfer inadvertently any residues from outside the flower into the nectar samples (see analytical report, page 24).

Sampling of Rape Flowers

About 10 g of rape flowers will be sampled from plants outside the tent area. After sampling, the flowers are stored on wet or on dry ice in the field. At the end of each sampling day at the latest, they were transferred into a refrigerator (-20°C) where they were retained until residue analysis (see 3.10).

3.10 Sample Processing and Residue Analysis

Sample processing and analytical methods are described in detail in appendix III.

3.11 Climatic Conditions During the Study

During the study, temperature and precipitation events were recorded two times. The following records were made:

Date	Precipitation	Min. temperature	Max. temperature	Remarks
16 June	4 mm	14°C	19°C	partly cloudy, windy
17 June	none	3°C	22°C	sunny, calm, dew formation in the morning
18 June	none	7°C	26°C	Sunny, calm, dew formation in the morning

3.12 Observations on Honeybees

During and after the test, study plots were examined for suspicious honeybee mortalities. In addition, the honeybees which were used as sampling devices were also observed for any signs of impairments. They were examined for the following behavioral impacts:

- exaggerated movements
- discoordinated movements
- apathy
- flight incapability

4.0 FILING

All raw data, the study protocol and the original of the report are filed in the Central GLP archive of PF/F, Crop Protection Center 40789 Monheim, FRG. Reserve samples of the test substance are stored in the pertinent archive of that test facility which provided or certified the test substance.

5.0 RESULTS AND DISCUSSION

Analytical findings are summarized in table 1 and given in detail in the analytical report (appendix I).

5.1 Analytical Findings

Analytical findings are summarized in table 1 and given in detail in the analytical report (appendix I). No residues at or above the limit of quantitation were found in any of the examined matrices for either the parent compound or the relevant metabolites (olefin- and hydroxy-imidacloprid.

5.2 Observations on Foraging Honeybees

No behavioral impacts (e.g. apathy, exaggerated motility, discoordinated movements) or suspicious mortality was observed on the honeybees used for collecting rape nectar and rape pollen. At the time of sampling, aphids were observed on the rape plants.

FIGURES

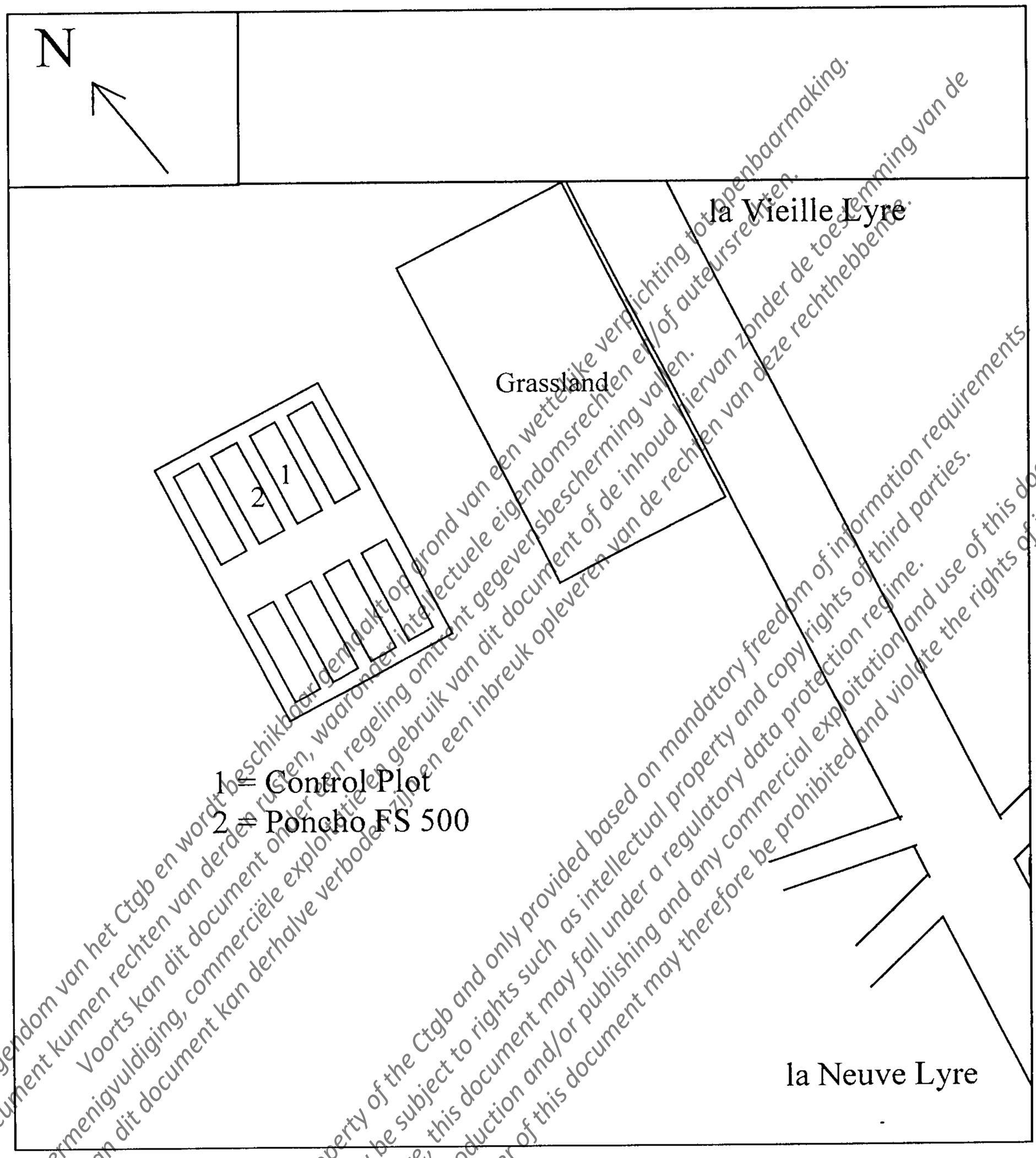


Figure 1: Location of the trial site and arrangement of the study plots.

There were two rows of 4 rape planted plots each on the trial site. Each plot had a size of 4x20 m with 1 m space between adjacent plots and 8 m space between the two plot rows. Crop management is reported under study no. RA-2042/98.

TABLES

Table 1: Summary of the Analytical Findings.

Type of Sample	Residue Level [mg/kg] *		
	Imidacloprid	Olefin-NTN	Hydroxy-NTN
Control Samples			
Honeybees before exposure	< 0.01	< 0.01	< 0.01
Rape nectar sampled by bees	--	--	--
Rape nectar sampled with micro-capillaries from the flowers	< 0.01	< 0.01	< 0.01
Rape blossoms	< 0.01	< 0.01	< 0.01
Rape pollen sampled by bees	--	--	--
Treatment Samples			
Honeybees before exposure	< 0.01	< 0.01	< 0.01
Rape nectar sampled by bees	< 0.01	< 0.01	< 0.01
Rape nectar sampled with micro-capillaries from the flowers	< 0.01	< 0.01	< 0.01
Rape blossoms	< 0.01	< 0.01	< 0.01
Rape pollen sampled by bees	< 0.01	< 0.01	< 0.01

* Limit of quantitation: 0.01 mg/kg

APPENDICES

APPENDIX I: Analytical Report.

[redacted]

Study No.: E 370 1358-7
(this appendix reports data from study no. RA-2042/98)

STUDY TITLE

Analysis of Rape Nectar, Rape Blossoms, and Rape Pollen for Residues of Imidacloprid and Imidacloprid Metabolites and Preliminary Observations of Effects on Domestic Honeybees in France

Residue Analytical Method for the Determination of Imidacloprid, Hydroxy- and Olefin-Metabolite in Rape Flower, Rape Pollen, Bee and Nectar Samples by HPLC-MS/MS

1) Nectar-Samples:

Extraction and sample clean-up:

1. Place for e.g. 1.0 g of the sample material in a 150-ml beaker.

2. Add 10 ml of water and place the sample for 2 min into a Ultrasonic Bath.

3. Add 20 ml of methanol.

4. Blend the sample using an ultra-turrax blender (or equivalent) for approximately 1 min.

5. Vacuum filter the suspension through 2.5 g of Celite filter aid using Schwarzband filter paper supported on a Büchner funnel into a 250-ml vacuum filter flask.

6. Wash the filtered solids with a total of 20 ml of methanol/water (3/1, v/v). Press residual solvent from the solids using rubber damming. Discard the filtered solids.

7. Transfer the filtrate into a 250-ml brown glass round-bottom flask.

8. Concentrate the filtrate to an aqueous remainder of 5 to 10 ml using a rotary evaporator with a max. bath temperature of 50 °C.

9. Add 5 to 10 ml water to the aqueous solution from step 8 to bring the total volume of the extracts to approx. 20 ml.

10. Place the aqueous solution on the top of the ChemElut® CE 1020 (20 ml volume) column fitted with a disposable stainless steel needle and wait for approx. 15 minutes to achieve an uniform distribution of the liquid on the column.

11. Elute the residues from the column with 140 ml of CH_2Cl_2. Collect the eluate in a 250-ml brown glass round-bottom flask.

12. Evaporate the eluate from step 11 to dryness using a vacuum rotary evaporator and a max. bath temperature of 40 °C.

13. Dissolve the residues in 1.00 ml of acetonitrile/water (2/8, v/v) and determine the residues with HPLC-MS/MS.

2.) Bee-Samples, Rape Flowers, Rape Pollen:

Extraction and sample clean-up:

1. Place for e.g. 2.0 g of the sample material in a 150-ml beaker. Add 30 ml of methanol/water (3/1,v/v) and allow the sample to soak for 30 min.

2. Blend the sample using an ultra-turrax blender (or equivalent) for approximately 1 min.

3. Vacuum filter the suspension through 2.5 g of Celite filter aid using Schwarzband filter paper supported on a Büchner funnel into a 250-ml vacuum filter flask.

4. Wash the filtered solids with a total of 30 ml of methanol/water (3/1, v/v). Press residual solvent from the solids using rubber damming. Discard the filtered solids.

5. Transfer the filtrate to a 100-ml graduated cylinder. Determine the total volume of the extracts. Mix the solution well, and transfer the half (e.g. 1.0 g sample equivalent) to a 250-ml brown glass round-bottom flask.

6. Concentrate the aliquot to an aqueous remainder of 5 to 10 ml using a rotary evaporator with a max. bath temperature of 50 °C.

7. Add 5 to 10 ml water to the aqueous solution from step 6 to bring the total volume of the extracts to approx. 20 ml.

8. Place the aqueous solution on the top of the ChemElut® CE 1020 (20 ml volume) column fitted with a disposable stainless steel needle and wait for approx. 15 minutes to achieve an uniform distribution of the liquid on the column.

9. Elute the residues from the column with 140 ml of CH_2Cl_2. Collect the eluate in a 250-ml brown glass round-bottom flask.

10. Evaporate the eluate from step 9 to dryness using a vacuum rotary evaporator and a max. bath temperature of 40 °C.

11. Dissolve the residues in 2 ml of toluene/ethyl acetate (85/15, v/v).

12. Apply the organic solution from step 11 onto a 0.5 g (3 ml) silica gel (SiOH) column (e.g. Varian).

13. Allow the solution to pass through the column at a flow rate of 1 ml/min.

14. Rinse the 250-ml brown glass round-bottom flask with 10 ml of toluene/ethyl acetate (70/30, v/v) and apply the solution onto the column, too.

15. Elute the residues with 5 ml of acetonitrile at a flow rate of 1 ml/min. Collect the eluate in a 25-ml brown glass pear-shaped flask.

16. Evaporate the eluate from step 15 to dryness using a vacuum rotary evaporator and a max. bath temperature of 40 °C. Dissolve the residues in e.g. 1.00 ml of acetonitrile/water (2/8, v/v) and determine the residues with HPLC-MS/MS.

<u>NOTE</u>

1. The volumes to be used for flushing the column with toluene/ethyl acetate and for elution with acetonitrile must be newly determined for each batch of SiOH-column!

2. The flow rate should not be too high, since otherwise losses of the residues in may occur with recoveries below 70 % and the clean-up is less effective.

3.) HPLC-MS/MS determination of Imidacloprid, Hydroxy and Olefin Metabolite:

A) Measuring equipment and HPLC conditions:

Instrument:	Hewlett Packard 1100
Column:	e.g.: Phenomenex, Luna C18 (2), 5 µm, 15 x 0.46 cm i.D. or Merck, Superspher, RP select-B, 4 µm, 12.5 x 0.4 cm i.D.
Solvent A:	Water + 0.1 ml Acetic acid/L
Solvent B:	ACN + 0.1 ml Acetic acid/L
Oventemperature:	40 °C
Inject.volume:	50 µL
Flow:	1.0 mL/min
Split:	150 µL into MS from 1000 µL

Time Table	0 min	20 % B
	10 min.	20 % B
	11 min	90 % B
	15 min	90 % B
	16 min	20 % B
	19 min	20 % B
	Stoptime	19 min

Retention Times:

Olefin-Imidacloprid	approx. 4.5 min
Hydroxy-Imidacloprid	approx. 5.5 min
Imidacloprid	approx. 8.5 min

B) *Mass Spectroscopy*

The experiments were performed on a triple-quadrupole mass spectrometer fitted with an electrospray interface operated in the positive ion mode under MRM conditions.

The mass spectrometer was tuned by infusing a standard solution of 0.5 mg/l Imidacloprid, Hydroxy-Metabolite and Olefin-Metabolite (dissolved in acetonitrile/water (2/8, v/v) + 0.1 ml acetic acid per litre) at a flow rate of 5-10 µl/min.

Mass axis calibration was done by infusing a polypropylene glycol 3000 solution. Unit mass resolution was established and maintained in each mass resolving quadrupole by maintaining a full widht at half-maximum of between 0.8 and 1.0 DA. After tuning and calibration, optimal collision-activated dissociation (CAD) conditions for fragmentation of Imidacloprid, Hydroxy-Metabolite and Olefin-Metabolite were determined. These experiments were performed with nitrogen as collision gas with a collision offset of -20 eV for Imidacloprid,

-23 eV for Hydroxy-Metabolite and -13 eV for Olefin-Metabolite at an approximate collision gas thickness of 1.56×10^{15} atoms/cm^2.

Nebulization gas is set at 1.48 l/min, curtain gas is set at 0.95 l/min and turbo gas is set at 6 l/min.

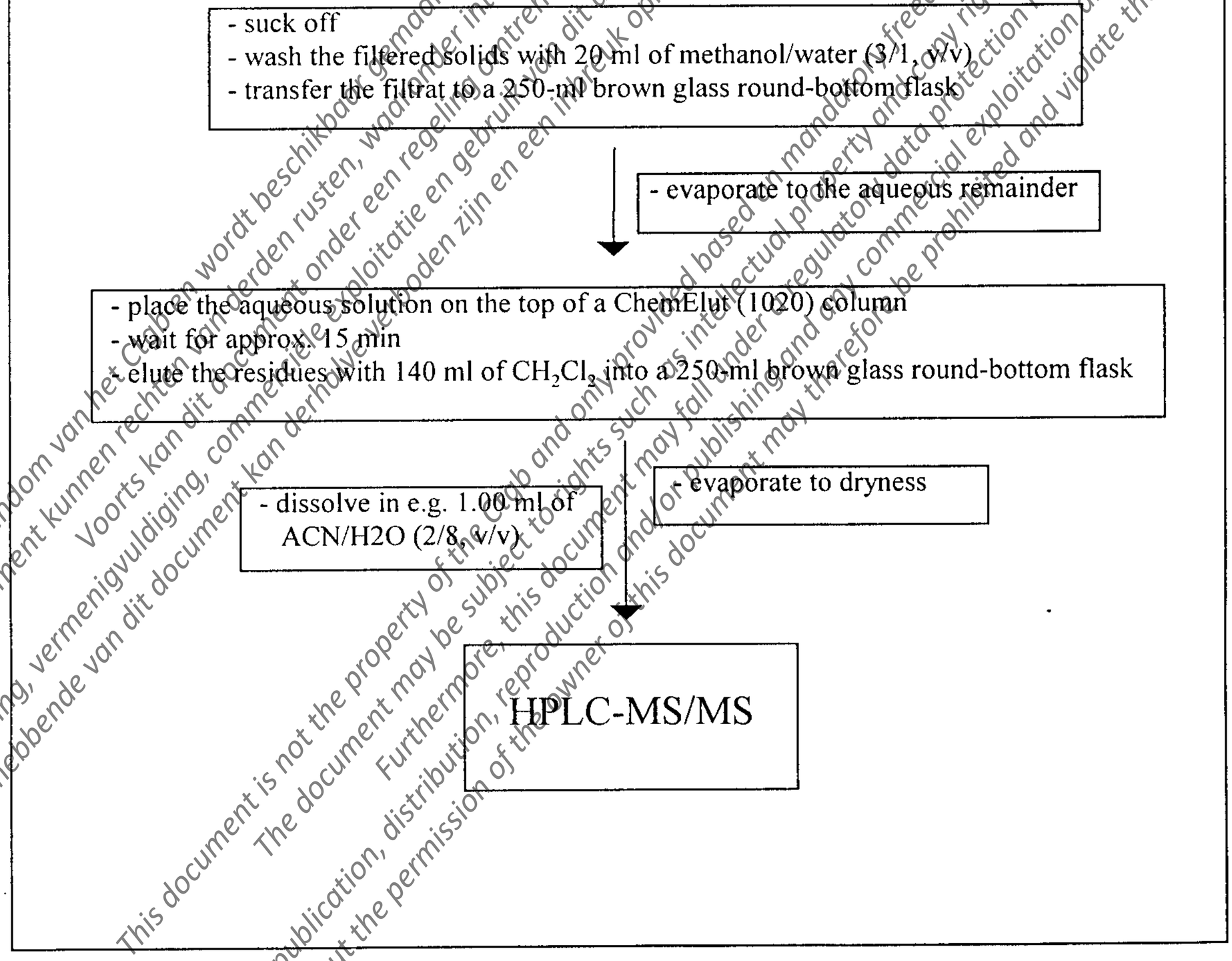
Detector:	e.g. Triple Quadrupol LC/MS/MS Mass Spectrometer, Perkin-Elmer Sciex Instruments API 300, Apple™ Macintosh® System 8.0
Interface:	Electrospray, TurboIon Spray Potential: +4900 V Temperature: 300 °C Nebulizer gas: Nitrogen 5.0 (99.999% purity), 1.48 l/min
Scan type:	MRM (Multiple Reaktion Monitoring Mode)
Polarity:	Positive
Aquisition mode:	Profile

Mass spectrometer operating parameters

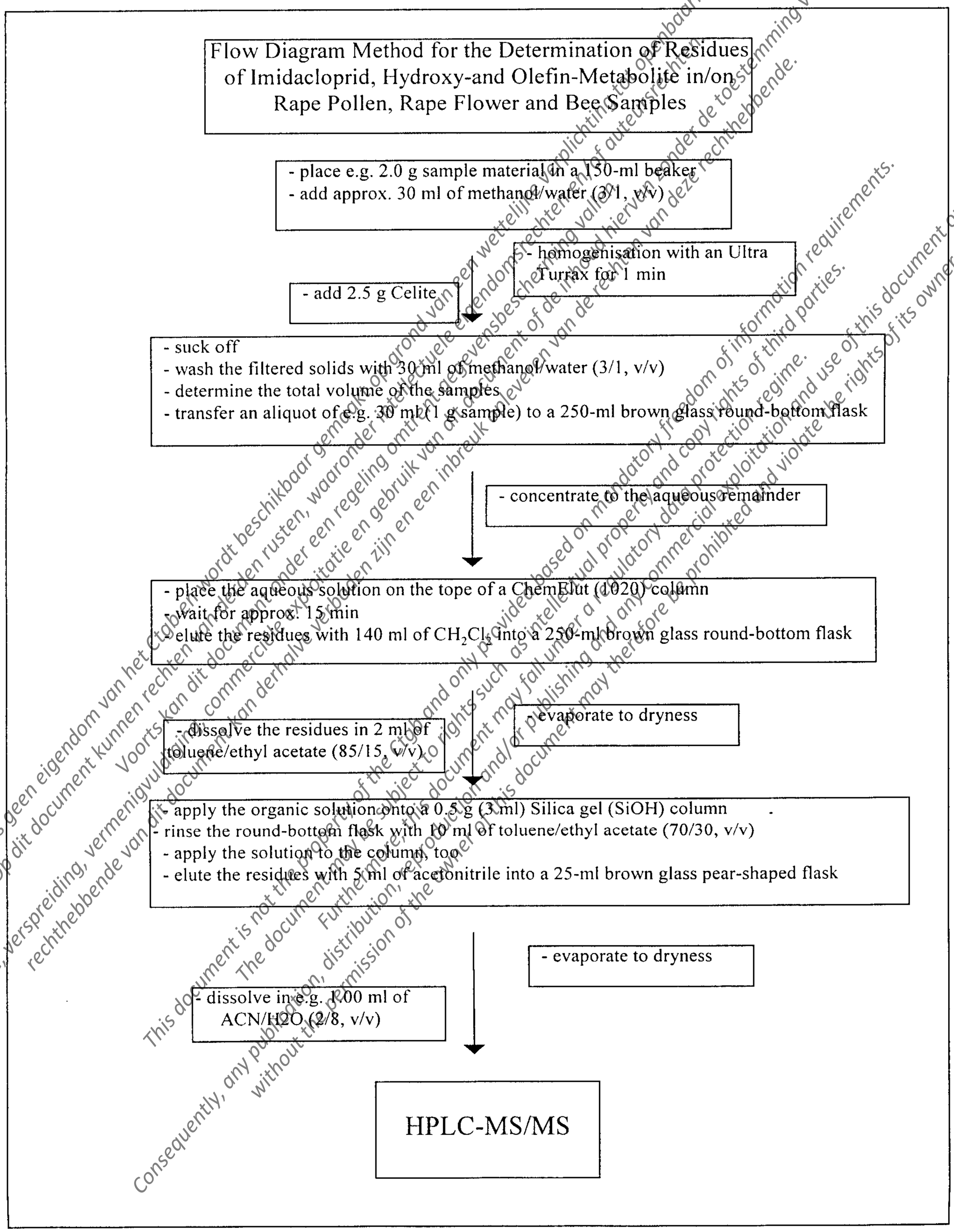
Compound	Precursor Ion Q1 Mass (amu)	Product Ion Q3 Mass (amu)	Dwell Time (msec)	Collision Energy (eV)
Imidacloprid (Cl 37)#	258.0	210.9	500	-20
Imidacloprid (Cl 35)	256.0	208.9	500	-20
Hydroxy-Metabolite (Cl 37)#	274.0	190.8	250	-23
Hydroxy-Metabolite (Cl 35)	272.0	190.8	250	-23
Olefin-Metabolite (Cl 37)#	256.0	237.8	250	-13
Olefin-Metabolite (Cl 35)	254.0	235.8	250	-13

#= ^{37}Cl isotope of all substances were detected to use as qualifiers

4.) Flow Diagram Method for the Determination of Residues of Imidacloprid and Metabolites in/on Nectar and Honey Samples



5.) Flow Diagram Method for the Determination of Residues of Imidacloprid and Metabolites in/on Rape Pollen, Rape Flower and Bee Samples:



6.) Results of Bee Samples, Nectar Samples of Bees, Rape Flower and Rape Pollen Samples.

A) Bee Samples:

Sample name	Sample description	Sample weight	Residues		
			Hydroxy-NTN [mg/kg]	Olefin-NTN [mg/kg]	Imidacloprid [mg/kg]
Samples taken before exposure					
Bees from hive 1 *	B Südfrankreich	19.6 g	< 0.01	< 0.01	< 0.01
Bees from hive 2 **	C Südfrankreich	21.2 g	< 0.01	< 0.01	< 0.01
Samples taken during exposure					
Bees Poncho	A Südfrankreich	8.8 g	< 0.01	< 0.01	< 0.01

* This hive was used to sample in the Poncho treated field

** This hive was used to sample in a field treated with a different compound. The results of this study are not reported here.

B) Nectar Samples:

Sample Name	Sample description	Sample weight	Residues		
			Hydroxy-NTN [mg/kg]	Olefin-NTN [mg/kg]	Imidacloprid [mg/kg]
Nectar harvested from the honeybulb of bees which were collected before exposure					
Bees from hive 1 *	B Südfrankreich N	1.769 g	< 0.01	< 0.01	< 0.01
Bees from hive 2 ** 1st sample	C Südfrankreich N	1.728 g	< 0.01	< 0.01	< 0.01
Bees from hive 2 ** 2nd sample	C Südfrankreich N II	1.401 g	< 0.01	< 0.01	< 0.01
Nectar harvested from the honeybulb of bees which were collected during exposure					
A Nectar N Poncho	A Südfrankreich N	1.287 g	< 0.01	< 0.01	< 0.01
Nectar harvested directly from the plants via micro-capillaries					
Nectar 06/16/98 Control	Nektar ca. 1 ml Eppendorfgefäß 16.06.98	ca.1 ml	< 0.01	< 0.01	< 0.01
Nectar 06/18/98 Control	Nektar ca. 1 ml Eppendorfgefäß 18.06.98	ca.1 ml	< 0.01	< 0.01	< 0.01
Nectar 06/16/98 Poncho	Nektar ca. 1 ml Eppendorfgefäß 16.06.98	ca.1 ml	< 0.01	< 0.01	< 0.01
Nectar 06/18/98 Poncho	Nektar ca. 1 ml Eppendorfgefäß 18.06.98	ca.1 ml	< 0.01	< 0.01	< 0.01

* This hive was used to sample in the Poncho treated field

** This hive was used to sample in a field treated with a different compound. The results of this study are not reported here.

C) Rape Flower Samples:

Sample Name	Sample description	Sample weight	Residues		
			Hydroxy-NTN [mg/kg]	Olefin-NTN [mg/kg]	Imidacloprid [mg/kg]
Rape Flower Control	Blüten in 2 Probengefäßen ca. 20 g 16.06.98	ca. 20 g	< 0.01	< 0.01	< 0.01
Rape Flower Poncho	Blüten in 2 Probengefäßen ca. 20 g 16.06.98	ca. 20 g	< 0.01	< 0.01	< 0.01

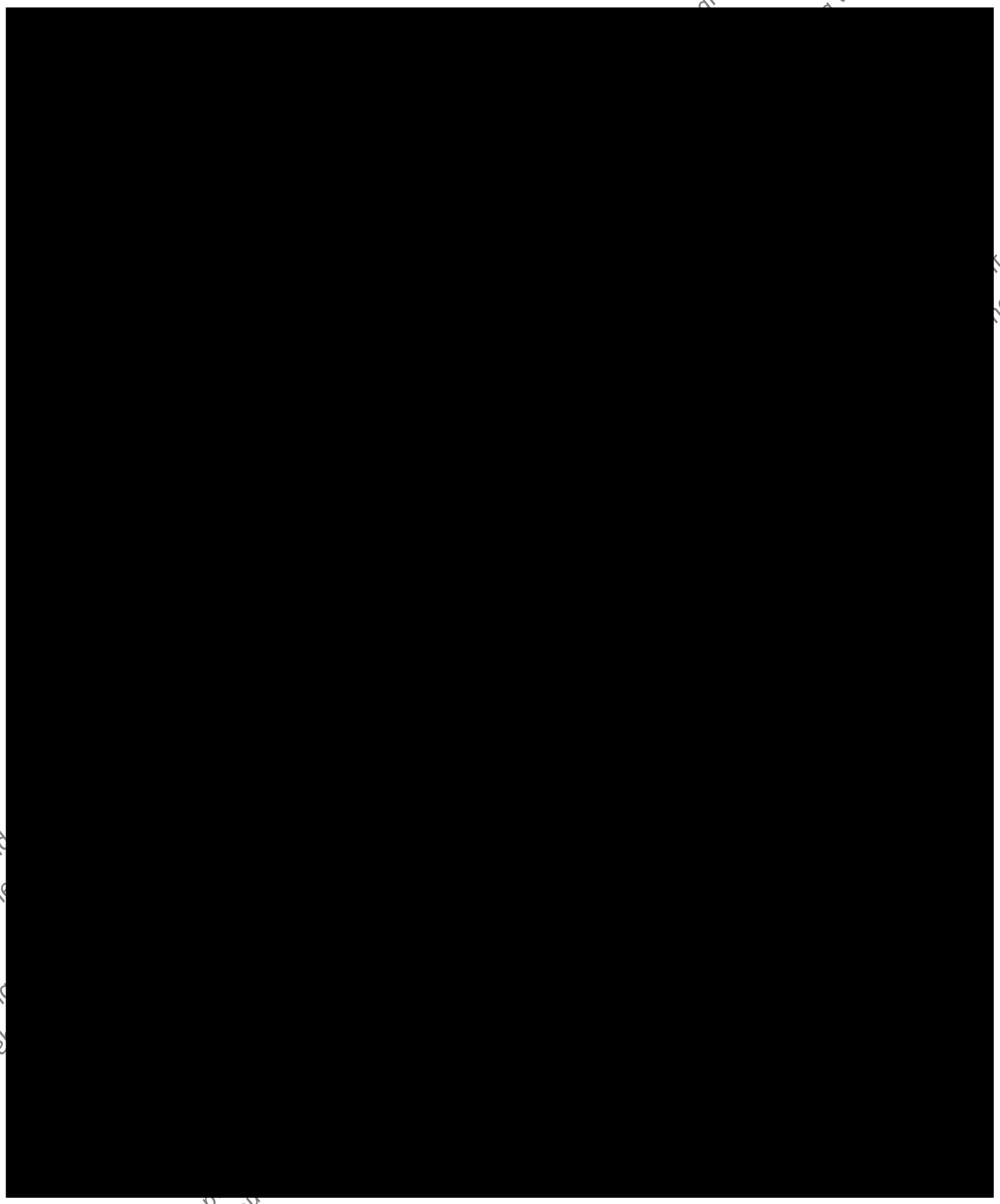
D) Rape Pollen Samples:

Sample Name	Sample description	Sample weight	Residues		
			Hydroxy-NTN [mg/kg]	Olefin-NTN [mg/kg]	Imidacloprid [mg/kg]
A Pollen Poncho	A Südfrankreich P	0.868 g	< 0.01	< 0.01	< 0.01

7.) Results of Water Control Samples

Sample Name	Sample description	Sample weight	Residues		
			Hydroxy-NTN [mg/kg]	Olefin-NTN [mg/kg]	Imidacloprid [mg/kg]
Water out of Plastic Bottle	Südfrankreich	1 g	< 0.01	< 0.01	< 0.01
Water out of Glas Bottle	Südfrankreich	1 g	< 0.01	< 0.01	< 0.01
Water FRA E 3701358-7 Blank Control Sample	Südfrankreich	1 g	< 0.01	< 0.01	< 0.01

Appendix II: Copy of the GLP Certificate



Appendix II (continued):

[redacted]

Appendix III: Quality Assurance Statement

Referat GLP
Quality Assurance Statement
[redacted]